



Optimization of experimental parameters for the determination of amoxicillin by sensitive spectrophotometric method using synthesized gold nanoparticles

Jafar Abolhasani^{1*}, Elham Motallebpour Sangestany¹, Behrouz Vahid²

1- Department of Chemistry, Tabriz branch, Islamic Azad University, Tabriz, Iran.

2- Department of Chemical Engineering, Tabriz Branch, Islamic Azad University, Tabriz, Iran

Received: 07/11/2013

Accepted: 15/12/2014

Published: 20/02/2014

Abstract

In this study, a sensitive and simple spectrophotometric method has been developed for determination of amoxicillin (AMX) using gold nanoparticles (AuNPs) synthesized by chemical reduction method. This method is based on addition of AMX to AuNPs and nanoparticles aggregation, consequently absorption band of nanoparticles decreased. The optimization of experimental variables was investigated by examining pH, temperature, buffer type and amount, AMX and AuNPs amounts and process time. In the desired condition, the linear calibration graph in the range of 10-250 µg/l was obtained with proper correlation coefficient ($R^2=0.99$). Limit of detection (LOD) and relative standard deviation (RSD %) were calculated as 0.12 µg/l and 0.61%, respectively. The proposed method was successfully applied to the AMX measurement in pharmaceutical, real water and biological samples.

Key words: Spectrophotometry, Amoxicillin, Gold Nanoparticles

1 Introduction

Amoxicillin (AMX) is one of the amino penicillin's classified as β -Lactam group [1, 2]. It is utilized widely in pharmaceutical industry for production drugs to treat various infections owing to its effectiveness, less side effects and low production cost [3]. Determination of AMX is significant for control of its production in biotechnological processes and also in the quality control of its preparation procedure [4]. Moreover, the presence of antibiotics in wastewater of hospital or pharmaceutical industries is a serious environmental threat due to the possibility of antibiotic-resistant strains formation in pathogenic bacteria [5]. Hence, monitoring of these species is very essential in aquatic environment [6]. However, AMX determination is not easy owing to the lack of any significant chromophores in its structure. Among various methods for determination of AMX, such as liquid chromatography-mass spectrometry [7,8], high performance liquid chromatography (HPLC) [9], high performance thin layer chromatography [10], derivative spectrophotometry [11,12], voltammetry [13], and capillary electrophoresis [14], HPLC is the most reported technique which needs large amount of pure solvents, derivatizing treatment and long equilibration time. Therefore,

application of a rapid, accurate, low cost and selective technique like spectrophotometric method for determination of AMX is essential from practical point of view [15]. Gold nanoparticles (AuNPs) have been applied widely and successfully for determination of pharmaceutical and biological compounds by varying of AuNPs absorption because of aggregation of nanoparticles in the presence of various analytes [16].

In this research work, simple and sensitive spectrophotometric process was developed for determination of AMX in aqueous solutions. First AuNPs were prepared with chemical reduction method [17]. Then, they were applied for determination of AMX amount by changing of experimental variables to find the optimal conditions. Eventually, results were used to find the AMX concentration in real samples including AMX capsule, spring water and biological samples.

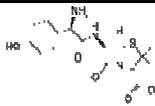
2 Materials and methods

2.1 Chemicals

Table 1 represents chemical structure and other characteristics of AMX, which is named chemically (2S, 5R, 6R)-6-[(R)-(-)-2-Amino-2-(p-hydroxyphenyl)acetamidol]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo [3.2.0] heptane-2-carboxylic acid trihydrate [18, 19].

Corresponding author: Jafar Abolhasani, Department of Chemistry, Tabriz branch, Islamic Azad University, Tabriz, Iran. E-mail: dj.abul@yahoo.com.

Table1: Characteristics of amoxicillin

| structure | Cass number | Molecular formula | λ_{\max} | Mw (g/mol) |
|---|-------------|-------------------------------------|------------------|------------|
|  | 26787-78-0 | $C_{16}H_{19}N_3O_5S_3 \cdot 3H_2O$ | 220 nm | 419.45 |

Standard reference of AMX and AMX capsules were provided from Dana Co. Iran. Other compounds which were used in this study purchased from Merck, Germany.

2.2 Synthesis of gold nanoparticles

Chemical reduction method was applied to prepare of AuNPs. Briefly, 250 ml of gold salt precursor (tetrachloroauric(III)acid trihydrate, 1.5 % v/v) was heated to 95 °C, then, 5 ml of trisodiumcitrate was added to the solution. Citrate is soluble in water acting as reluctant agent of Au(III). It has negative charge preventing aggregation AuNPs in solution. The formation of pink solution with maximum absorbance λ_{\max} at 520 nm confirmed the production of AuNPs [20, 21].

2.3 Experimental procedure

All experiments were performed in a batch mode. A solution with certain amount of AMX and AuNPs (A), (by pH, buffer type, amount of gold nanoparticles, amount of buffer, temperature, and process time) was prepared with total volume of 3ml by addition of double distilled water. Then, the absorbance of each solution was recorded against the blank solvent (A_0) and ($A-A_0$) determined at λ_{\max} of 520 nm. The pH of solution was measured by pH meter (pH211, Hanna, Romany) and hydrochloric acid (0.1 M) or sodium hydroxide (0.1 M). The effect of different buffers at The influence of various volume of acetate buffer in the range of 200-800 μ l was investigated. As can be seen in Fig.2, addition of 200 μ l of acetate buffer solution resulted in maximum absorption of AuNPs.

3.1.1 Effect of pH

The influence of pH on the absorbance of AuNPs was investigated in the range of 3-9 (Fig.1). The maximum absorption was observed at pH of 5. In pHs less than 5, AuNPs aggregated and the absorption declined. Above desired pH the absorption decreased because of deprotonation of amin group of AMX and more aggregation of AuNPs [22]. Place the cursor at the beginning of the first line in either of the columns and press the required button.

3.1.2 Effect of buffer types during experiment

pH stabilization at optimum value is essential in determination of AMX. In order to study the role of buffer, acetate and phosphate buffers was selected with concentration of 0.5M in the same experimental variables with Fig 2. Results revealed that AuNPs had more absorption using of acetate buffer (0.61) in comparison of phosphate (0.42) due to closer pKa to optimum pH [23].

3.1.3 Effect of buffer volume

The influence of various volume of acetate buffer in the range of 200-800 μ l was investigated. As can be seen in Fig.2, addition of 200 μ l of acetate buffer solution resulted in maximum absorption of AuNPs.

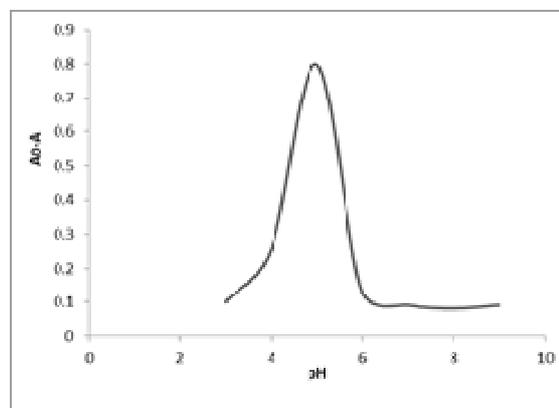


Fig. 1. Effect of pH on AuNPs absorbance at $\lambda_{\max} = 520$ nm, AMX = 60 μ g/L, AuNPs = 1000 μ l and T = 25°C

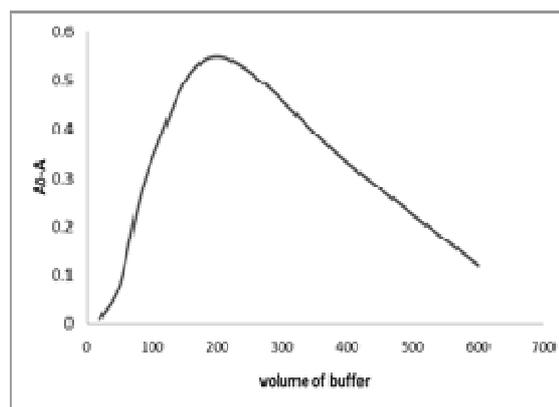


Fig.2. Effect of buffer volume on AuNPs absorbance at $\lambda_{\max} = 520$ nm, AMX = 60 μ g/l, AuNPs = 1000 μ l, buffer volume = 200 μ l and T = 25 °C

3.1.4 Effect of AuNPs volume

The AuNPs amount in the solution is key parameter for determination of AMX. Hence, the influence of AuNPs amount was investigated by varying AuNPs volume in the range of 200-900 μ l (Fig. 3). Addition of 700 μ l of AuNPs to the solution led to desired absorption. Below 700 μ l there were not adequate AuNPs in the solution and above this value, more aggregation took place resulting in less absorption and sensitivity [24].

3.1.5 Effect of temperature and process time

Process time and temperature showed negligible influence on absorbance of AuNPs at the same experimental variables for determination of AMX. Moreover, the product of AuNPs with AMX was stabilized instantly at 25°C and remained stable after 30h.

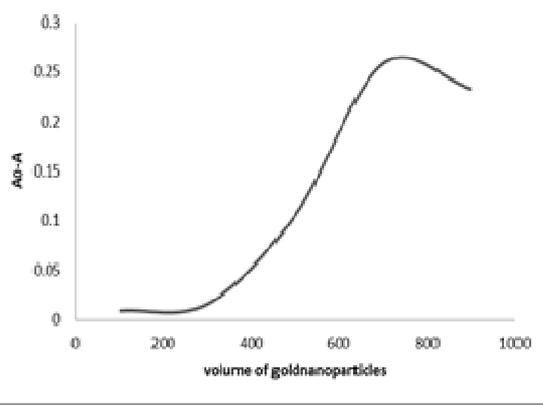


Fig. 3. Effect of AuNPs volume on AuNPs absorbance at λ_{max} = 520 nm. AMX = 60 μ g/L, AuNPs = 700 μ l, buffer volume = 200 μ l and T = 25 $^{\circ}$ C

3.1.5 Effect of amoxicillin concentration

The absorbance of AMX with different concentrations at optimal conditions was demonstrated in Fig. 4. The absorbance declined with increasing of AMX amounts due to more aggregation of AuNPs in the presence of drug [25].

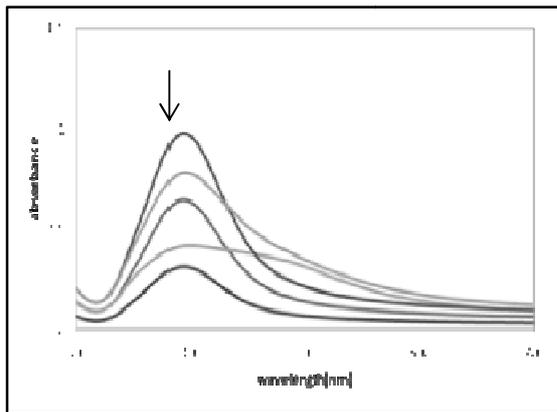


Fig. 4. The UV-Vis absorption spectra of AuNPs in the absence and presence of AMX with concentrations of 10 μ l , 100 μ l ,200 μ l and 250 μ l . AMX = 60 μ g/L, AuNPs = 700 μ l, buffer volume = 200 μ l and T = 25 $^{\circ}$ C

3.2 Validation

3.2.1 Calibration graph

After selecting desired experimental variables the calibration graph was plotted at AMX concentration range of 10-250 μ g/l (Fig.5). The obtained straight line with high correlation coefficient of 0.99 which was obtained based on bear-lambert law indicated appropriate linearity.

3.2.2 Limit of detection and precision

The limit of detection (LOD) was calculated by Eq. (1) as 0.21.

$$LOD=(3 S_0)/b \tag{1}$$

Where S_0 is standard deviation of blank solution and b is the slope of calibration graph. The precision of the suggested method was evaluated by relative standard

division at optimized conditions (Eq. (2)) for AMX amount of 60 μ g/l, which was repeated for six times as 0.61 % .

$$RSD\%=(S \times 100)/m \tag{2}$$

Where S and m are standard deviation and average of AuNPs absorbance, respectively.

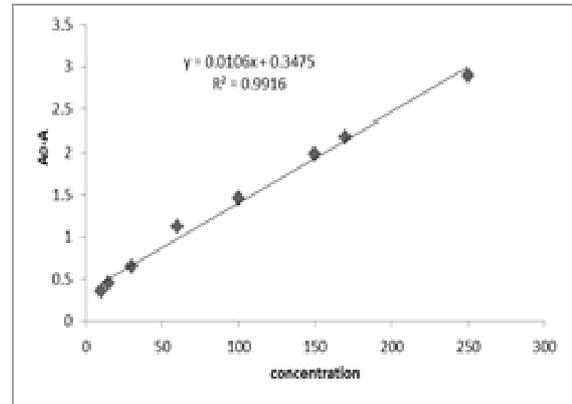


Fig. 5. AuNPs calibration graph in presence of AMX at λ_{max} = 520 nm. AMX = 60 μ g/L, AuNPs = 700 μ l, buffer volume = 200 μ l and T = 25 $^{\circ}$ C

3.2.3 Specificity

The determination of 60 μ g/l of AMX was studied in presence of various foreign species. Table 2 demonstrates the maximum tolerance value of the investigated species, which was determined when absorbance value did not exceed \pm 5% on addition of them.

Table 2: Determination of AMX in presence of various foreign species at λ_{max} = 520 nm.

| Sample No. | Foreign species | Maximum tolerance limit (μ g/l) |
|------------|---|--------------------------------------|
| 1 | Mg ²⁺ , Ca ²⁺ , k ⁺ , Cl ⁻ , Ni ²⁺ , Na ⁺ | 1000 \leq |
| 2 | Co ²⁺ , Ba ²⁺ , Sr ²⁺ | 500 |
| 3 | I ⁻ | 250 |
| 4 | EDTA | 100 |
| 5 | Bi ³⁺ | 10 |
| 6 | Cu ²⁺ , Fe ²⁺ | 1 |

AuNPs = 700 μ l , acetate buffer = 200 μ l , pH = 5, AMX = 60 μ g/l and T = 25 $^{\circ}$ C

3.3 Analytical application

To investigate the ability of the proposed method for measuring the drug in real samples, it was utilized to determine AMX in pharmaceutical, spring water and biological samples. Recovery experiments for spiked samples solution were performed for evaluation of the method. Recovery ranged from 99.8-100.4% revealing adequate accuracy, which was calculated by Eq. (3). The obtained data presented in Table 3.

$$Recovery \% = (m-c)/spiked \times 100 \tag{3}$$

Table 3: Real sample recovery at λ_{max} = 520 nm

| sample | Added amount | Amax | Recovery % |
|------------------------------|--------------|-------|------------|
| Capsule | 10 μ l | 1.064 | 99.8% |
| Spring water (sardrood area) | 10 μ l | 1.062 | 97.5% |
| Biological sample (urine) | 10 μ l | 1.01 | 99.3% |

AuNPs = 700 μ l , acetate buffer = 200 μ l , pH = 5, AMX = 60 μ g/l and T = 25 $^{\circ}$ C

4 Conclusions

In this research, synthesized gold nanoparticles with chemical reduction method were utilized for determination of amoxicillin by the sensitive spectrophotometric method. This method was based on the aggregation of gold nanoparticles as a consequence of amoxicillin addition to them; hence, nanoparticles absorption declined. After optimization of experimental variables including pH, temperature, buffer type and amount, AMX and AuNPs amounts and process time, calibration plot was depicted at $\lambda_{\text{max}} = 520\text{nm}$ in the range of 10-250 $\mu\text{g/l}$ with adequate correlation coefficient of 0.99. Limit of detection and relative standard deviation of proposed method were calculated as 0.12 $\mu\text{g/l}$ and 0.61%, respectively. Amoxicillin amount could be successfully determined in pharmaceutical, real water and biological samples.

References

- 1- R, Ding., P.zhang.,M.seredych.,T.bandosz.(2011).. Removal of antibiotics from water using sewage sludge- and waste oil sludge derived adsorbents. *Water Research*46(13): 4081-4090.
- 2- Xu, Wei-hai., Zhang, Gan.,Zou, Shi-chun.,Li, Xiangdong .,Liu, Yu-chun. (2007), Determination of selected antibiotics in the Victoria Harbour and the Pearl River, South China using high-performance liquid chromatography-electrospray ionization tandem mass spectrometry. *Environmental Pollution*. 145(3): 672-679.
- 3- R, Cogo., G ,Caiazzo., P, De Luca1., V, Boddi. (1994), Corresponding author contact information, Massimo De Luca3, Alessandro Casini3, Effect of miocamycin and amoxicillin/clavulanate on total serum immunoglobulin E levels in patients with infectious exacerbations of allergic asthma: A crossover trial." *Current Therapeutic Research* 55(2): 184-198.
- 4- K ,Srirangan, et al(2013).. Biotechnological advances on Penicillin G acylase: Pharmaceutical implications, unique expression mechanism and production strategies.. *Biotechnology Advances* 31(8): 1319-1332.
- 5- P ,Liu (2014).. Removal of trace antibiotics from wastewater: A systematic study of nanofiltration combined with ozone-based advanced oxidation processes.. *Chemical Engineering Journal*240(0): 211-220.
- 6- F. I ,Khattab, Safa'a, M., Riad, Mamdouh ,R., Rezk, Mohamed K., Abd El-Rahman, Hoda., M Marzouk(2013), A single novel PVC membrane for dual determination of sulphadimethoxine and malachite green in aquatic environment.. *Arabian Journal of Chemistry*.
- 7- Hui, Li., Xi, Xia.,YananXue., Shusheng ,Tang., Xilong ,Xiao., Jiancheng, Li (2012).. Simultaneous determination of amoxicillin and prednisolone in bovine milk using ultra-high performance liquid chromatography tandem mass spectrometry.. *Journal of Chromatography B* 900(0): 59-63.
- 8- ChuangjiLiu., Hai ,Wangb., Yanbin, Jiangb., Zhenxia ,Du (2011).. Rapid and simultaneous determination of amoxicillin, penicillin G, and their major metabolites in bovine milk by ultra-high-performance liquid chromatography-tandem mass spectrometry.. *Journal of Chromatography B* 879(7-8): 533-540.
- 9- J.I.D, Wibawa., D, Fowkes., P,N Shaw., D,A Barrett (2012).. Measurement of amoxicillin in plasma and gastric samples using high-performance liquid chromatography with fluorimetric detection.. *Journal of Chromatography B* 774(2): 141-148.
- 10- A.M, Idris and Elgorashe, E. E (2011).. Sequential injection chromatography against HPLC and CE: Application to separation and quantification of amoxicillin and clavulanic acid." *Microchemical Journal* 99(2): 174-179.
- 11- C,Bosch Ojeda, and Sanchez Rojas, F(2013).. Recent applications in derivative ultraviolet/visible absorption spectrophotometry: 2009-2011: A review. *Microchemical Journal* 106(0): 1-16.
- 12- G.G,Mohamed(2001).. Spectrophotometric determination of ampicillin, diclucaxillin, flucloxacillin and amoxicillin antibiotic drugs: ion-pair formation with molybdenum and thiocyanate. *Journal of Pharmaceutical and Biomedical Analysis* 24(4): 561-567.
- 13- R, Ojani.,J,Raoof., S, Zamani (2012).. A novel voltammetric sensor for amoxicillin based on nickel-curcumin complex modified carbon paste electrode. *Bioelectrochemistry* 85(0): 44-49.
- 14- Deng, B., Shi ,A., Li, L., Kang,Y (2008).. Pharmacokinetics of amoxicillin in human urine using online coupled capillary electrophoresis with electrogeneratedchemiluminescence detection." *Journal of Pharmaceutical and Biomedical Analysis* 48(4): 1249-1253.
- 15- Q-Q,Liang, and Li,Y.-S. A(2013).. rapid and accurate method for determining protein content in dairy products based on asynchronous-injection alternating merging zone flow-injection spectrophotometry. *Food Chemistry* 141(3): 2479-2485.
- 16- C, Lee., M, A., G, Alison., A, Weiss., P, Zhang(2013).. Colorimetric viral detection based on sialic acid stabilized goldnanoparticles." *Biosensors and Bioelectronics* 42(0): 236-241.
- 17- I.O, Ali(2013).. Synthesis and characterization of Ag0/PVA nanoparticles via photo- and chemical reduction methods for antibacterial study. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 436(0): 922-929.
- 18- M. Marques, C. Raposo, M. Damasceno, M. Filho, D. Pinto, and E. Barroso(2006).. Simultaneous determination of amoxicillin and Cefalexin in human plasma by LC-MS/MS. 17 IMSC, Prague.
- 19- S. Bogialli, V. Capitolino, R. Curini, A. Corcia, M. Nazzari and M. Sergi(2013).. Simple and rapid liquid chromatography-tandem mass spectrometry confirmatory assay for determining amoxicillin and ampicillin in Bovine tissue and milk. *J. Agric. Food Chem* 52(11): 3286-91.
- 20- B-H,Sohn, B-W,Seo, S-I,Yoo(2002).. Changes of the Lamellar PeriodbyGoldNanoparticles in the Nanoreactor Scheme of Thin Films of Symmetric Diblock Copolymers. *J. Mater. Chem*, 12, 1730-1734.

- 21- J. Wang, T. Zhu, M. Tang, S. M. Cai and Z. F. Liu(1996),. Fabricating Surface Enhanced Raman Scattering (SERS)-Active Substrates by Assembling Colloidal Au Nanoparticles with Self-Assembled Monolayers. *Jpn. J. Appl. Phys.* 35
- 22- M. Dousa, R. Hosmanova(2005),. *Journal of Pharmaceutical and Biomedical Analysis*, 37 373.
- 23- H.T.S. Britton(1942),. Chapter XVI: Solutions of known hydrogen ion concentration In: *Hydrogen Ions*; vol. I, Chapman and Hall Ltd.: London. p. 304.
- 24- M.A ,Metrick(2013),. The effects of buffers and pH on the thermal stability, unfolding and substrate binding of RecA. *Biophysical Chemistry* 184 (0): 29-36.
- 26- B, Gilbert. Reyn ,K. Ono, Kristen. A, Ching(2009),. The effects of nanoparticle aggregation processes on aggregate structureand metal uptake. *Journal of Colloid and Interface Science* 339 285–295.